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Impact of Physical Activity In the Prevention of Colorectal Cancer

Sarah Ashley Barnes
University of South Carolina

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IMPACT OF PHYSICAL ACTIVITY IN THE PREVENTION OF COLORECTAL CANCER

by

Sarah A Barnes

Bachelor of Science
Newberry College, 2008

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School of Medicine

University of South Carolina

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Accepted by:

Dr. Angela Murphy, Director of Thesis

Dr. Walden Ai, Reader

Dr. Edie Goldsmith, Reader

Lacy Ford, Vice Provost and Dean of Graduate Studies

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DEDICATION

To my trainees: may each workout bring you closer to health and happiness.

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First and foremost, I'd like to thank my mentor, Dr. Angela Murphy for her positive role and unending supportive patience throughout the writing process.

I'd also like to thank Dr. Edie Goldsmith and Dr. Walden Ai for their contributions to my thesis work. Lastly, I could not have sat still long enough to complete this work had my boyfriend Wayne Fisher not lovingly, but persistently, encouraged me to do so. Thank you Love.

ABSTRACT

This review evaluates the current understanding of research on the impact of physical activity in the prevention of colorectal cancer. Current biological mechanisms implicated in physical activity and colorectal cancer risk reduction are blood glucose regulation, insulin sensitivity, leptin and adiponectin profiles, inflammation as well as secreted protein acidic and rich in cysteine (SPARC), an exercise induced myokine. Recent literature indicates that 30-60 minutes of moderate to vigorous activity a day is effective against colorectal cancer development, and there is convincing evidence of aerobic exercise as differently beneficial in recruiting mechanisms identified as preventative against colorectal cancer. This article provides a critical review of the evidence-based literature concerning the benefits of physical activity in reducing the risk for colorectal cancer. Further well designed animal and clinical trials testing differing exercise protocols are recommended for future research to enable better understanding of the currently implicated mechanisms in colorectal cancer development.

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LIST OF ABBREVIATIONS

WHO.....	World Health Organization
AOM.....	Azoxymethane
DMH.....	1,2-Dimethyl Hydrazine
SPARC.....	Secreted Protein Acidic and Rich in Cystine
IGF-I.....	Insulin Growth Factor 1
IGF-II.....	Insulin Growth Factor 2
IGFBP-3.....	Insulin Growth Factor Binding Protein 3
IGFBP-6.....	Insulin Growth Factor Binding Protein 6
DSS.....	Dextran Sodium Sulphate
IMCE.....	Immortomouse Colon Epithelial Cells
APN.....	Adiponectin
MCP-1.....	Monocyte Chemoattractant Protein 1
BMI.....	Body Mass Index
PA.....	Physical Activity
REE.....	Resting Energy Expenditure

CHAPTER 1

INTRODUCTION

According to the American Cancer Society's 2013 statistics, colorectal cancer is the third most prevalent cancer among men and women in the United States. Despite mortality rates declining in the population over 50 years of age, rates of colorectal cancer incidence in individuals under 50 years of age are on the rise. The etiology of colorectal cancer is a complex phenomenon that involves the interaction of genetic and environmental factors. However, the vast majority of cases can be ascribed to environmental causes as they account for more than 80% of all incidences ¹. Physical inactivity has emerged as a leading environmental risk factor for colorectal cancer. According to the World Health Organization (WHO), physical inactivity is the fourth leading risk for mortality globally; it is responsible for 6% of all deaths worldwide. In fact, physical inactivity raises the risk for numerous chronic diseases such as heart disease ²⁻⁶, diabetes ⁷⁻¹¹ and various cancers including colorectal cancer ¹²⁻¹⁷. Physical inactivity is thought to account for 13-14% of all colorectal cancer cases ¹⁸. Thus, the American Cancer Society speculates that the increase in colorectal cancer incidence among the younger population is, at least in part, due to a sedentary lifestyle. Conversely, physical activity or exercise training has been linked to a reduced risk for colorectal cancer; epidemiological studies indicate an inverse association between physical activity and colorectal cancer risk and controlled experiments in mice substantiate these claims. While there is an abundance of data to support a link between physical activity and

colorectal cancer risk, the mechanisms responsible for this relationship have not yet been fully elucidated.

Possible mechanisms that link physical activity to reduced colorectal cancer risk have been investigated using epidemiological studies as well as controlled experimental studies in mice. Mouse models that recreate the colorectal cancer development seen in humans have been particularly useful in examining the benefits of exercise on colorectal cancer risk in a controlled setting (i.e the dose of exercise can be controlled). For example, the $Apc^{Min/+}$ mouse that develops intestinal colon polyps similar to an aggressive case of human colorectal cancer has been widely used in exercise studies. Also, carcinogens that induce colorectal cancer including azoxymethane, (AOM) and 1,2-dimethyl hydrazine, (DMH) have been utilized to investigate the effects of exercise on colorectal cancer in both mice and rats. To date, the mechanisms that have been implicated in the benefits of physical activity on colorectal cancer risk include insulin regulation, leptin and adiponectin profiles, inflammatory processes as well as secreted protein acidic and rich in cysteine (SPARC), an exercise induced myokine. This review will examine the recent literature on the aforementioned mechanisms that are thought to link physical activity to reduced colorectal cancer risk.

CHAPTER 2

METABOLIC MEDIATORS

Metabolic mediators control growth and development, regulate energy output and storage, and work to maintain healthy body mass. Imbalances in their concentrations have been linked to a heightened risk for colorectal cancer.

2.1 INSULIN

Insulin is a hormone produced by beta cells in the pancreas; it functions to regulate blood glucose levels. Evidence indicates that insulin resistance and its associated pathological conditions including elevated fasting glucose and insulin are implicated in colorectal cancer¹⁹. Risk factors for colorectal cancer, including obesity, physical inactivity and type 2 diabetes are all linked to insulin resistance and hyperinsulinemia^{20,21,22,23,24}. While epidemiological evidence only indirectly associates insulin with colorectal cancer risk, strong evidence implicates insulin as player in colorectal cancer. For example, insulin injections have been reported to enhance the growth of aberrant crypt foci as well as increase the number and size of colorectal tumors^{25,26}. Similarly, *in vivo* evidence supports a tumor promoting role of insulin on colorectal cancer cells^{27,28,29,30}. Alterations in insulin may play a role in the benefits of exercise on the development of colorectal cancer. It is well recognized that exercise can decrease hyperinsulinemia. For example, in a study of one hundred Type 2 diabetic obese or overweight subjects, exercise was successful at reducing plasma insulin levels.

Within this high-risk population, a significant decrease in plasma insulin levels across all exercise modalities was reported. The greatest effect was seen in those subjects performing aerobic training in conjunction with resistance training³¹. To date, there is no direct evidence linking exercise to the prevention of colorectal cancer via a reduction in insulin levels. However, lowering baseline serum levels of insulin through exercise would presumably attenuate insulin's well-documented effects on colorectal cancer development.

2.2 INSULIN GROWTH FACTOR 1

Insulin-like growth factors (IGF-I and IGF-II) and their binding proteins (IGFBP-1 and IGFBP-6) have been linked to colorectal cancer risk in epidemiological studies as well as controlled experimental studies in mice^{32,33}. IGFs promote proliferation and survival of a wide range of cell types and are thus critical for normal development. However, dysregulation of the IGF system has been implicated in a range of diseases including colorectal cancer. For example, epidemiological studies have shown that higher circulating IGF levels are associated with increased risk for colorectal cancer³⁴. This relationship is supported by controlled experimental studies in mice; colorectal cancer growth is decreased in mice following deletion of the IGF1 gene³⁵. *In vitro* studies have documented that the IGF-1 receptor facilitates malignant transformation of a number of oncogenes³⁶. And there is an abundance of literature to support the role of IGF-1 on promotion of survival, proliferation, migration and invasion of many cell types. For example, *in vitro* IGF-1 promotes cell cycle progression and inhibition of apoptosis in several colonic adenocarcinoma cell lines^{37,29}.

Exercise training has been shown to influence IGF levels, which may be a potential mechanism for the benefits of exercise on colorectal cancer. However, the dynamics of this relationship is likely heavily influenced by the intensity, mode and duration of the training regime. Several studies suggest that initiation of a training program, which is associated with a substantial increase in energy expenditure, leads initially to decreases in IGF levels. Conversely, following adaptation to the training the suppression of IGF is diminished and in fact IGF may exceed the pre-training levels. Therefore, the relationship between exercise and IGF is potentially very complex, which likely explains the inconsistent findings on its potential in the literature. For example, one study reported that voluntary wheel running exercise was associated with a lower polyp number in *Apc^{min}* mice but the exercise group actually showed higher IGF-1 levels; although there was no association between polyp number and IGF reported ³⁸. Contrary to those findings, recent literature has shown that IGF-1 levels are slightly lowered (~9%) a carcinogenic mouse model (AOM/DSS) of colorectal cancer following voluntary exercise ³⁹. However, the decrease was only significantly correlated with the lower colon tumor formation in AOM/DSS-treated mice when the associated increasing IGFBP-3 levels were considered ³⁹. Ju *et al* hypothesized a lower molar ratio of IGF-1 to IGFBP - 3 (IGF binding protein) was responsible for inhibiting tumor growth in these mice ³⁹. Although studies in animal literature support a positive effect of exercise on IGF-1 in colorectal cancer, the evidence has been mixed, possibly due to the complex nature of IGF-1's interactions with a number of other contributing factors as well as the diversity of exercise protocols used in these studies. More research detailing IGF-1's responses to exercise are needed to formulate a better evidence base within the mouse model. To date,

there are no clinical studies on the effects of exercise on IGF-1 in colorectal cancer, this is likely due, at least in part, to the conflicting evidence foundation in the animal literature.

CHAPTER 3

ADIPOKINES

Adipose-derived hormones (adipokines), along with their customary role in energy homeostasis, have been identified as potential mediators of the effects of exercise on colorectal cancer risk.

3.1 LEPTIN

Leptin is a long-term regulator of energy expenditure and food intake. Leptin has been shown to stimulate intestinal oncogenesis through a variety of signaling pathways⁴⁰. The hypothesized link between leptin and colorectal cancer was driven initially by reports of leptin receptor expression by various human epithelial colon cancer cell lines. When bound to leptin, the receptor can activate signal transduction pathways that can enhance cell proliferation and DNA synthesis. For example, Fenton et al., reported that leptin promotes the proliferation of preneoplastic (IMCE) cells⁴¹. While the *in vitro* evidence is relatively strong, there are mixed reports among the animal studies. A recent study found a significant decrease of tumor cell proliferation in leptin-deficient tumors, as well as a dramatic inhibition of tumor growth in leptin-deficient and leptin-receptor-deficient mice⁴². Whereas a previous study reported that leptin failed to promote growth of colon cancer xenografts in nude mice and did not increase intestinal tumorigenesis in *Apc^{Min/+}* mice⁴³. Clinical investigations have reported that a progressive increase in leptin occurs during colorectal carcinogenesis⁴⁴. Taken together, the available literature suggests that lowering baseline levels of leptin could lower risk for developing colorectal cancer. It

has been shown that exercise can significantly reduce baseline leptin levels in APC^{Min} mice⁴⁵. However, it is important to note that while small changes were seen in male polyp development, no decrease in female mice tumor load was observed; therefore, the relevance of these findings are inconclusive. Follow up studies by Colbert *et al* demonstrated that a greater frequency and duration of voluntary wheel running decreased male polyp count by 25%, but in those studies, energy balance, rather than leptin was the mechanism investigated³⁸. Citing Colbert *et al*'s decision to conclude investigating leptin mechanistically after their first study, it can be presumed that although dated animal research found that exercise was effective in lowering baseline leptin levels, current research does not support a leptin mediated link between exercise and colorectal cancer prevention. However, it is important to note that the observed decrease in leptin with exercise, independent of colorectal cancer, is likely due to the exercise-induced decrease in fat mass.

3.2 ADIPONECTIN

Adiponectin (APN) is an adipokine secreted exclusively by adipose tissue, which has been reported to be negatively associated with colorectal cancer risk; a decrease in adiponectin is associated with increased risk for colorectal cancer. Adiponectin's suppressive role in tumorigenesis has been widely documented in mouse models of colorectal cancer^{46,47}. More recent animal studies have used APN deficiency models to mechanistically determine its role in both the risk and development of colorectal cancer. For example, Saxena *et al* reported that APN deficiency was associated with higher expressions of pro-inflammatory and pro-cancerous markers⁴⁸. Further, APN knockout mice showed greater tumor numbers as well as greater tumor area due to an increase in

colonic mucosal erosion that resulted from a thinning of the mucosal lining⁴⁹. Another study examined the role of APN in a carcinogenic as well as a genetic mouse model of colorectal cancer. It was found that deficiency of APN increased tumorigenesis in both models; in the *Apc*^{min/+} mouse model APN^{-/-} increased tumorigenesis by 3.2 fold and in the carcinogenic model by ~70%⁵⁰.

Several studies have examined the benefits of exercise on APN in the settings of colorectal cancer. To date, Saxena *et al* has reported the most promising data to support the protective effects of exercise on colorectal cancer development. They report increased serum levels of APN by exercise training and suggest that it may be one of the mechanisms for the protective role of exercise of inflammatory processes in a carcinogenic mouse model of colitis.⁵¹ As expected, their APN deficient mice were more susceptible to inflammation than their wild-type counterparts and the benefits of exercise on inflammation were lost in their APN deficient mice. Other studies have demonstrated similar effects of exercise on APN levels. Tang *et al* noted a particularly interesting shift of APN levels in rats in response to exercise. High fat diet feedings for 8 weeks decreased circulating APN levels; however, 8 weeks of exercise training reversed this effect⁵². Such results prove promising for prompting additional studies to improve our understanding of the effects of exercise on adiponectin in the settings of colorectal cancer.

As with the animal literature, higher levels of APN have been implicated in having a protective role in the development of human colorectal cancer. For example, an inverse relationship between serum APN and colorectal cancer grade was observed in a recent case study involving subjects diagnosed with colorectal cancer⁵³. Further, the

benefits of exercise on APN have also been investigated, although not in the settings of colorectal cancer. To date, few clinical trials have investigated the specific effects of exercise on APN levels. This is presumably due to the fact that the effect of exercise on circulating APN levels varies by individual; this may be attributed to variances in genes or environmental factors ⁵⁴. A study by Lee *et al* sought to understand the activity of APN polymorphisms and completed a study on obese women having variations of two polymorphisms of the APN gene that are reported to be associated with insulin resistance and APN levels ⁵⁵. They found that regardless of polymorphism, aerobic training over 3 months increased circulating APN levels ⁵⁶. Because studies in mouse models have reported that an increase in APN levels decreases polyp development and load, presumably these subjects would have attenuated risk for colorectal cancer given the exercise-induced increase in APN. In addition, these individuals had a significant decrease in body weight, BMI, and waist circumference, which would also mitigate their risk of developing colorectal cancer. Although the clinical literature available seems promising, further clinical studies are needed to confirm the link between exercise, adiponectin and the development of colorectal cancer outside the animal model.

CHAPTER 4

INFLAMMATION

The colon has evolutionarily outfit itself with a large population of immunologically active cells to defend the body against invasion and to regulate homeostasis ⁵⁷. It is through their modulation that the release of inflammatory cytokines is regulated ⁵⁸. If, over time, these cells are constantly expressing higher than normal levels of these inflammatory cytokines it can lead to chronic intestinal inflammation. It has been hypothesized not only that colorectal cancer pathogenesis may be influenced by the state of this chronic intestinal inflammation ⁵⁹, but also that regular exercise is capable of mitigating it.

4.1 TNF ALPHA

TNF- α is a cytokine involved primarily with systemic inflammation. Its effector cells (immune cells) regulate the inflammatory response to a number of different assaults and through initiation of a variety of inflammatory cascades. TNF- α has been found to be a main stimulant of colitis-associated colorectal cancer ⁵⁹. The most likely mechanism for this response is that TNF- α elevates the activity of inflammation-sensitive transcription factors such as NF- κ B ⁶⁰ as well as the inflammatory cytokines IL-6 and IL-1 β ⁶¹. We have shown that TNF- α mRNA expression is increased in the intestines of the Apc^{Min/+} mouse, which is associated with the abundance of large polyps ⁶². Further, using a monocyte chemoattractant protein 1 (MCP-1) knockout mouse we have reported a decrease in the number of macrophages in the polyp tissue, which was linked to a

reduction in TNF- α expression ⁶³. This is of particular importance given that the predominant expression of TNF- α in colorectal cancer is observed within tumor associated macrophages ⁶⁴. Direct evidence for the involvement of TNF- α in cancer comes from observations that antibodies against TNF- α inhibit the development of inflammation-related colon cancer ⁶⁵.

While the available literature on the benefits of exercise on TNF- α in colon cancer is limited, the studies that are presented are promising. For example, recently it has been shown that regular exercise suppressed the generation of aberrant crypt foci in the colon following AOM and this was associated with decreased levels of TNF- α in both the colon and plasma ⁶⁶. Similarly, TNF- α expression in intestinal lymphocytes of younger C57BL/6 mice has been shown to be decreased with voluntary wheel running exercise suggesting that long term exercise may protect the bowel by reducing intestinal inflammation ⁶⁷. Further, a recent study in aged C57BL/6 mice demonstrated that regular exercise lowered TNF- α in plasma and serum ⁶⁸. Although good evidence for a benefit of exercise on TNF- α in colorectal cancer in animals is beginning to accumulate in the literature, the clinical data is disappointingly sparse and relatively undocumented. None-the-less a team in India has demonstrated that humans respond accordingly to regular exercise, with over 70% of subjects lowering their baseline values of TNF-alpha after one month of moderate exercise ⁶⁹.

4.2 IL 6

Interleukin-6 or (IL)-6 is a pleiotropic cytokine that modulates a variety of physiological responses and activates genes associated with cellular proliferation, differentiation, and apoptosis. Epidemiological studies indicate an association between

IL-6 and colorectal cancer and controlled experiments in mice substantiate these claims. This effect is thought to be largely mediated through its ability to increase intestinal inflammation ⁷⁰. A recent study reported that IL-6 expression in stage III colon cancer patients is a prognostic marker of tumor behavior ⁷¹. Further, using IL-6 knockout mice as well as IL-6 overexpression techniques, Baltgalvis *et al.*, eloquently examined the role of IL-6 on tumor characteristics in the $Apc^{Min/+}$ mouse model of colon cancer. They reported that knocking out IL-6 reduced the number of large polyps by approximately 30% and these effects were reversed when IL-6 was overexpressed ⁷². Interestingly, they also demonstrate that IL-6 is necessary for the onset of adipose and skeletal muscle wasting in the $Apc^{Min/+}$ mouse as $IL-6^{-/-}$ mice did not lose gastrocnemius muscle mass or epididymal fat pad mass while IL-6 overexpression led to a decrease in mass for each of these tissues.

IL-6 has been implicated in the link between exercise and colorectal cancer; several studies have reported a benefit of exercise on IL-6 in colorectal cancer. One study reported that nine weeks of moderate treadmill running decreased total intestinal polyps by 29% in male $Apc^{Min/+}$ mouse and this was associated with a 98% decrease in IL-6 ⁷³. A follow up study by the same group documented that moderate-intensity treadmill exercise can attenuate IL-6-dependent cachexia in $Apc^{Min/+}$ mice, independent of changes in IL-6 concentration and muscle inflammatory signaling ⁷⁴. The exercise effect was associated with improved insulin sensitivity and improved energy status in the muscle. Similarly, IL-6 induced mitochondrial remodeling and proteolysis was documented to be rescued with moderate exercise training even in the presence of high circulating IL-6 levels ⁷⁵. While there is good evidence to support an effect of exercise

on IL-6 in mouse models of colorectal cancer, there is very little clinical evidence in this area. Although given the well documented effects of moderate exercise training on reducing IL-6, it is likely that these same outcomes will be seen in the human literature.

CHAPTER 5

SPARC

It has been postulated that the exercise-induced production of novel myokines can alter the mechanisms involved in colorectal cancer pathogenesis. In a recent study, Aoi *et al.* suggested that the anti-tumor effect of regular exercise was more dependent on circulating factors rather than endogenous proteins in the colon ⁶⁶. One such myokine, secreted protein acidic and rich in cysteine (SPARC) also known as osteonectin, is a matricellular protein that is involved in cell to cell interaction, growth factor function and cell differentiation ⁷⁶. A recent study by Aoi *et al* examined the effects of exercise on the expression of this novel myokine in the muscle of mice and humans following a single bout of exercise. It was found that a single bout of aerobic exercise increased plasma levels of SPARC in both humans and mice via promotion of SPARC upregulation in contracting myocytes ⁷⁷. Although SPARC returned to baseline levels after 6 hours in humans, their studies demonstrate that regular exercise enhanced the secretory ability of SPARC in response to muscle contraction by increasing the amount of SPARC in resting muscle tissue ⁷⁷. Using SPARC knockout mice, they went on to examine if the benefits of exercise on colorectal cancer may be mediated by SPARC. They found that low-intensity exercise reduced aberrant crypt foci growth in a chemically-induced mouse model of colorectal cancer. However, the benefits of exercise were lost in SPARC deficient mice implicating a role of SPARC on the benefits of exercise in colorectal cancer. Further, exercise enhanced apoptosis in colon mucosal cells but again these

effects were not evident in SPARC deficient mice. These recent findings concerning up-regulation of SPARC via aerobic exercise provide yet another outlet to investigate the promising effects of exercise in prevention of colorectal cancer. Further studies are needed to better elucidate SPARC's effects on the known mechanisms of colorectal pathogenesis.

CHAPTER 6

PHYSICAL ACTIVITY AND EXERCISE RECOMMENDATIONS

Physical activity (PA) is defined by the American College of Sports Medicine as any bodily movement created from muscle contraction and resulting in an energy expenditure increase over that of resting energy expenditure (REE). Exercise is a planned and structured form of PA that uses repetitive bodily movement to improve or maintain one or more components of physical fitness, which are simply characteristics that allow one to perform physical activities.

The World Health Organization's Global Recommendations on Physical Activity for Health (2008) focus mainly on type, duration, frequency, intensity, and volume of physical activity. These recommendations are minimum guidelines allotted to provide benefits in cardiorespiratory health, metabolic health, bone health, depression and breast and colon cancer. WHO recommends at least 150 minutes of moderate intensity aerobic physical activity or at least 75 minutes of vigorous physical activity per week. WHO's guidelines also note that at least 30-60 minutes per day of moderate to vigorous activity is needed to observe significantly lower risks of breast and colon cancer⁷⁸.

While epidemiological data suggested higher durations of physical activity is protective against colorectal cancer, the early animal literature was inconclusive. Colbert *et al* noted that while prior epidemiological studies and studies in chemically induced cancerous rats concluded a beneficial influence of exercise on tumor polyp load, their 2002 work provided limited evidence for possible preventative effects against intestinal

tumorigenesis in male $Apc^{min/+}$ mice, and no evidence for an effect in female $Apc^{min/+}$ mice ⁴⁵. Subsequently, advances in exercise protocols as well as additional investigation into pivotal mechanisms surrounding the development of colorectal cancer have established a dose-response relationship supporting the suppressive nature of exercise on colorectal cancer.

Recent studies by Colbert *et al* with C57BL/6J- Apc^{Min} mice reported that voluntary treadmill running and the negative energy balance resulting from pair-feeding with a non-exercising control group decreased polyp number in a dose dependent manner in the running group. The running group also maintained a significantly smaller overall body weight than the control group ³⁸. Similarly, another study reported that forced low-intensity running and forced swimming reduced aberrant crypt foci, precursors to colonic adenocarcinomas, sustained from carcinogenic DMH-injections in rats ^{79,80}. More recently, in mice subjected to azoxymethane (AOM) injections, a tri-weekly running schedule for six weeks decreased aberrant crypt foci development compared to a sedentary treatment ⁶⁶.

While there is ample evidence to support a lower incidence of colorectal cancer development in animal models with higher levels of aerobic exercise, the clinical data supporting such a link is scarce by comparison. However, McTiernan *et al*, found a 12-month exercise intervention of 60 minutes per day 6 days a week decreased colonic crypt cell proliferation in healthy human subjects ⁸¹. Because an increased cell proliferation rate and an extension of the normal proliferative zone within the colon crypt are reversible precursors of colonic neoplasia ⁸², these subjects, having lowered their proliferation rates, would have attenuated their risk for developing colorectal cancer.

Likewise, a 2011 meta-analysis of 20 clinical studies of physical activity and colonic polyps by Wolin *et al* concluded a significant inverse relation between physical activity and colon adenomas or polyps. Wolin *et al* conclude that physical activity may reduce the risk of colon polyp development by 15%. The reduction of risk by physical activity may also be substantially greater for large and advanced polyps⁸³. Recent cohort studies also mirror animal and clinical findings on the positive role of physical activity on colon cancer risk. A 2012 screening study of 912 multi-ethnic persons scheduled for an exam concluded that more than 60 minutes of exercise a week, was correlated with a low or no polyp count. Subjects who performed at least 60 minutes of exercise weekly were less likely to have any detectable polyps compared to those who did not regularly exercise. They were also less likely to have an adenoma on their colonoscopy compared to their counterparts with more sedentary lifestyles⁸⁴. The questions in the study pertaining to exercise were asked in a manner that inferred activity outside daily activity, i.e. exercise. Unfortunately, mode of exercise was not investigated in this study.

CHAPTER 7

CONCLUSION

The current body of evidence supports a positive effect of exercise, and in particular vigorous aerobic activity, on the prevention of colorectal cancer. Animal studies support the hypothesis that exercise mediates this protective effect by inducing changes in blood glucose regulation, insulin sensitivity, the adiponectin profile, inflammation regulation and SPARC secretion. For a better understanding of these mechanisms and their role in preventing tumorigenesis, further studies containing even more exercise protocol variations should be completed. Granted the scarcity of current clinical data, future clinical studies are needed to support the conclusions of these animal studies. Further advancements within this field are important for public health prevention practices and awareness that may aid in nudging the population towards healthier lifestyle choices.

REFERENCES

1. Rustgi AK. The genetics of hereditary colon cancer. *Genes Dev* 2007;21:2525-38.
2. Mendis S. The contribution of the Framingham Heart Study to the prevention of cardiovascular disease: a global perspective. *Prog Cardiovasc Dis*;53:10-4.
3. Carnethon MR. Physical Activity and Cardiovascular Disease: How Much is Enough? *Am J Lifestyle Med* 2009;3:44S-9S.
4. Wise FM. Coronary heart disease--the benefits of exercise. *Aust Fam Physician*;39:129-33.
5. Schwandt P, Liepold E, Bertsch T, Haas GM. Lifestyle, Cardiovascular Drugs and Risk Factors in Younger and Elder Adults: The PEP Family Heart Study. *Int J Prev Med*;1:56-61.
6. Arsenault BJ, Rana JS, Lemieux I, et al. Physical inactivity, abdominal obesity and risk of coronary heart disease in apparently healthy men and women. *Int J Obes (Lond)*;34:340-7.
7. Alibegovic AC, Sonne MP, Hojbjerre L, et al. Insulin resistance induced by physical inactivity is associated with multiple transcriptional changes in skeletal muscle in young men. *Am J Physiol Endocrinol Metab*;299:E752-63.
8. Thyfault JP, Booth FW. Lack of regular physical exercise or too much inactivity. *Curr Opin Clin Nutr Metab Care*;14:374-8.
9. Admiraal WM, van Valkengoed IG, JS LdM, Stronks K, Hoekstra JB, Holleman F. The association of physical inactivity with Type 2 diabetes among different ethnic groups. *Diabet Med*;28:668-72.
10. Eckardt K, Taube A, Eckel J. Obesity-associated insulin resistance in skeletal muscle: role of lipid accumulation and physical inactivity. *Rev Endocr Metab Disord*;12:163-72.
11. Hojbjerre L, Sonne MP, Alibegovic AC, et al. Impact of physical inactivity on subcutaneous adipose tissue metabolism in healthy young male offspring of patients with type 2 diabetes. *Diabetes*;59:2790-8.

12. Friedenreich CM, Neilson HK, Lynch BM. State of the epidemiological evidence on physical activity and cancer prevention. *Eur J Cancer*;46:2593-604.
13. Joshi CE, Parmigiani G, Colditz G, Platz EA. Opportunities for the primary prevention of colorectal cancer in the United States. *Cancer Prev Res (Phila)*;1:138-45.
14. Nahleh Z, Bhatti NS, Mal M. How to reduce your cancer risk: mechanisms and myths. *Int J Gen Med*;4:277-87.
15. Wiggins MS, Simonavice EM. Cancer prevention, aerobic capacity, and physical functioning in survivors related to physical activity: a recent review. *Cancer Manag Res*;2:157-64.
16. Ott JJ, Ullrich A, Mascarenhas M, Stevens GA. Global cancer incidence and mortality caused by behavior and infection. *J Public Health (Oxf)*;33:223-33.
17. Kokkinos P, Sheriff H, Kheirbek R. Physical inactivity and mortality risk. *Cardiol Res Pract*;2011:924945.
18. Harriss DJ, Cable NT, George K, Reilly T, Renehan AG, Haboubi N. Physical activity before and after diagnosis of colorectal cancer: disease risk, clinical outcomes, response pathways and biomarkers. *Sports Med*;37(11):947-60.
19. Giovannucci E. Insulin and colon cancer. *Cancer Causes Control*;6:164-179.
20. DeFronzo RA, Ferrannini E. Insulin resistance: a multifaceted syndrome responsible for NIDDM, obesity, hypertension, dyslipidemia, and atherosclerotic cardiovascular disease. *Diabetes Care* ;14:173-194.
21. Bardou M, Barkun AN, Martel M. Obesity and colorectal cancer. *Gut*. 2013 Mar 12.
22. Yoon J, Son K, Eom C, Durrance D, Park S. Pre-existing diabetes mellitus increases the risk of gastric cancer: A meta-analysis. *World J Gastroenterol*; 19(6):-936-945.
23. Khalili H, Chan AT. Is diabetes a risk factor for colorectal cancer? *Dig Dis Sci*;57(6):1427-9.
24. Kim YI. Diet, lifestyle, and colorectal cancer: is hyperinsulinemia the missing link. *Nutr Rev*;56:275-279.
25. Corpet DE, Jacquinet C, Peiffer G, et al. Insulin injections promote the growth of aberrant crypt foci in the colon of rats. *Nutr Cancer*;27:316-320.

26. Koohestani N, Tran TT, Lee W, et al. Insulin resistance and promotion of aberrant crypt foci in the colons of rats on a high fat diet. *Nutr Cancer*;29:69–76.
27. Pollak M. Insulin and insulin-like growth factor signaling in neoplasia. *Nat Rev Cancer*. 2008;8:915–928.
28. Pollak M. Insulin, insulin-like growth factors and neoplasia. *Best Pract Res Clin Endocrinol Metab*. 2008;22:625–638.
29. Guo YS, Narayan S, Yallampalli C, et al. Characterization of insulin like growth factor I receptors in human colon cancer. *Gastroenterology*. 1992;102:1101–1108.
30. Singh P, Guo YS, Narayan S, et al. IGF-I and IGF-I receptor in mouse colon. *In Vitro Cell Dev Biol*;27A(10):755-8.
31. Kadoglou NP, Fotiadis G, Athanasiadou Z, Vitta I, Lampropoulos S, Vrabas IS. The effects of resistance training on ApoB/ApoA-I ratio, Lp(a) and inflammatory markers in patients with type 2 diabetes. *Endocrine*;42(3):561-9.
32. Ma J, Pollak MN, Giovannucci E, Chan JM, Tao Y, Hennekens CH, Stampfer MJ. Prospective study of colorectal cancer risk in men and plasma levels of insulin-like growth factor (IGF)-I and IGF-binding protein-3. *J Natl Cancer Inst*.1999Apr 7;91(7):620-5.
33. Manousos O, Souglakos J, Bosetti C, Tzonou A, Chatzidakis V, Trichopoulos D, Adami HO, Mantzoros C. IGF-I and IGF-II in relation to colorectal cancer. *Int J Cancer*. 1999 Sep 24;83(1):15-7.
34. Chi F, Wu R, Zeng YC, Xing R, Liu Y. Circulation insulin-like growth factor peptides and colorectal cancer risk: an updated systematic review and meta-analysis. *Mol Biol Rep*. 2013 May;40(5):3583-90.
35. Wu, Y. P., Yakar, S., Zhao, L., Hennighausen, L. and LeRoith, D. Circulating insulin-like growth factor-I levels regulate colon cancer growth and metastasis. *Cancer* 2002, Res. 62,1030–1035
36. Baserga, R., Peruzzi, F. and Reiss, K. The IGF-1 receptor in cancer biology. *Int. J. Cancer* 2003, 107, 873–877
37. Koenuma M, Yamori T, Tsuruo T (1989) Insulin and insulin-like growth factor 1 stimulate proliferation of metastatic variants of colon carcinoma 26. *Jpn J Cancer Res* 80:51–58

38. Colbert LH, Mai V, Tooze JA, Perkins SN, Berrigan D, Hursting SD. Negative energy balance induced by voluntary wheel running inhibits polyp development in APCMin mice. *Carcinogenesis*. 2006 Oct;27(10):2103-7.
39. Ju J, Nolan B, Cheh M, Bose M, Lin Y, et al. Voluntary exercise inhibits intestinal tumorigenesis in ApcMin/+ mice and azoxymethane/dextran sulfate sodium-treated mice. *BMC Cancer*. 2008;8: 316.
40. Sikalidis AK, Varamini B. Roles of hormones and signaling molecules in describing the relationship between obesity and colon cancer. *Pathol Oncol Res*. 2011 Dec;17(4):785-90.
41. Fenton JI, Hursting SD, Perkins SN, Hord NG. Leptin induces an Apc genotype-associated colon epithelial cell chemokine production pattern associated with macrophage chemotaxis and activation. *Carcinogenesis* 2007;28:455-64.
42. Endo H, Hosono K, Uchiyama T, et al. Leptin acts as a growth factor for colorectal tumours at stages subsequent to tumour initiation in murine colon carcinogenesis. *Gut*;60:1363-71.
43. Aparicio T, Kotelevets L, Tsocas A, et al. Leptin stimulates the proliferation of human colon cancer cells in vitro but does not promote the growth of colon cancer xenografts in nude mice or intestinal tumorigenesis in Apc(Min/+) mice. *Gut* 2005;54:1136-45.
44. Mariusz Koda, Mariola Sulkowska, Luiza Kanczuga-Koda, Eva Surmacz, and Stanislaw Sulkowski. Overexpression of the obesity hormone leptin in human colorectal cancer. *J Clin Pathol*.2007 August;60(8): 902–906.
45. Colbert LH, Mai V, Perkins SN, Berrigan D, Lavigne JA, Wimbrow HH, Alvord WG, Haines DC, Srinivas P, Hursting SD. Exercise and intestinal polyp development in APCMin mice. *Med Sci Sports Exerc*.2003 Oct;35(10):1662-9.
46. Otani K, Kitayama J, Yasuda K, Nio Y, Iwabu M, Okudaira S, Aoki J, Yamauchi T, Kadowaki T, Nagawa H. Adiponectin suppresses tumorigenesis in Apc(Min)(/+) mice. *Cancer Lett*. 2010 Feb 28;288(2):177-82.
47. Fenton JI, Birmingham JM, Hursting SD, Hord NG. Adiponectin blocks multiple signaling cascades associated with leptin-induced cell proliferation in Apc Min/+ colon epithelial cells. *Int J Cancer*;122(11):2437-45.
48. Saxena A, Chumanovich A, Fletcher E, Larsen B, Lattwein K, Kaur K, Fayad R. Adiponectin deficiency: role in chronic inflammation induced colon cancer. *Biochim Biophys Acta*. 2012 Apr;1822(4):527-36.

49. Saxena A, Baliga M, et al. Mucus and adiponectin deficiency: role in chronic inflammation-induced colon cancer. *Int J Colorectal Dis.* 2013 Mar 9.
50. Mutoh M, Teraoka N, Takasu S, Takahashi M, et al. Loss of Adiponectin Promotes Intestinal Carcinogenesis in Min and Wild-type Mice. *Gastroenterology.* 2011 Jun;140(7):2000-8.
51. Saxena A, Fletcher E, Larsen B, Baliga MS, Durstine JL, Fayad R. Effect of exercise on chemically-induced colitis in adiponectin deficient mice. *J Inflamm (Lond).* 2012 Aug 21;9(1):30.
52. Tang H, Xie M, et al. The roles of aerobic exercise training and suppression IL-6 gene expression by RNA interference in the development of insulin resistance. *Cytokine.*;61(2):394-405.
53. Gialamas SP, Petridou ET, Tseleni-Balafouta S, Spyridopoulos TN, Matsoukis IL, Kondi-Pafiti A, Zografos G, Mantzoros CS. Serum adiponectin levels and tissue expression of adiponectin receptors are associated with risk, stage, and grade of colorectal cancer. *Metabolism.*;60(11):1530-8.
54. Woo SK, Kang HS. Apolipoprotein C-III SstI genotypes modulate exercise-induced hypotriglyceridemia. *Med Sci Sports Exerc.* 2004 Jun;36(6):955-9.
55. González-Sánchez JL, Zabena CA, Martínez-Larrad MT, Fernández-Pérez C, Pérez-Barba M, Laakso M, Serrano-Ríos M. An SNP in the adiponectin gene is associated with decreased serum adiponectin levels and risk for impaired glucose tolerance. *Obes Res*;13(5):807-12.
56. Lee KY, Kang HS, Shin YA. Exercise improves adiponectin concentrations irrespective of the adiponectin gene polymorphisms SNP45 and the SNP276 in obese Korean women. *Gene*;516(2):271-6.
57. Lefrançois L, Lycke N. Isolation of mouse small intestinal intraepithelial lymphocytes, Peyer's patch, and lamina propria cells. *Clin Chem*; 52(1):97-103.
58. Powrie F. Immune regulation in the intestine: a balancing act between effector and regulatory T cell responses. *Ann N Y Acad Sci*;1029:132-41.
59. Terzić J, Grivennikov S, Karin E, Karin M. Inflammation and colon cancer. *Gastroenterology*;138(6):2101-2114.
60. Greten FR, Eckmann L, Greten TF, Park JM, Li ZW, Egan LJ, Kagnoff MF, Karin M. IKKbeta links inflammation and tumorigenesis in a mouse model of colitis-associated cancer. *Cell*;118(3):285-96.

61. Wilson JA. Tumor necrosis factor alpha and colitis-associated colon cancer. *N Engl J Med*;358(25):2733-4.
62. McClellan JL, Davis JM, Steiner JL, Day SD, Steck SE, Carmichael MD, Murphy EA. Intestinal inflammatory cytokine response in relation to tumorigenesis in the *Apc(Min/+)* mouse. *Cytokine*;57(1):113-9.
63. McClellan JL, Davis JM, Steiner JL, Enos RT, Jung SH, Carson JA, Pena MM, Carnevale KA, Berger FG, Murphy EA. Linking tumor-associated macrophages, inflammation, and intestinal tumorigenesis: role of MCP-1. *Am J Physiol Gastrointest Liver Physiol*;303(10):G1087-95.
64. Naylor MS, Stamp GW, Balkwill FR. Investigation of cytokine gene expression in human colorectal cancer. *Cancer Res*; 50 (14):4436-4440.
65. Onizawa M, Nagaishi T, Kanai T, Nagano K, Oshima S, Nemoto Y, Yoshioka A, Totsuka T, Okamoto R, Nakamura T, Sakamoto N, Tsuchiya K, Aoki K, Ohya K, Yagita H, Watanabe H. Signaling pathway via TNF-alpha/NF-kappaB in intestinal epithelial cells may be directly involved in colitis-associated carcinogenesis. *Am. J. Physiol. Gastrointest. Liver Physiol.*;296 (4):G850-859
66. Aoi W, Naito Y, Takagi T, Kokura S, Mizushima K, Takanami Y, Kawai Y, Tanimura Y, Hung LP, Koyama R, Ichikawa H, Yoshikawa T. Regular exercise reduces colon tumorigenesis associated with suppression of iNOS. *Biochem Biophys Res Commun.*;399(1):14-9.
67. Hoffman-Goetz L, Pervaiz N, Guan J. Voluntary exercise training in mice increases the expression of antioxidant enzymes and decreases the expression of TNF-alpha in intestinal lymphocytes *Brain Behav Immun*;23(4):498-506.
68. Packer N, Hoffman-Goetz L. Exercise training reduces inflammatory mediators in the intestinal tract of healthy older adult mice. *Can J Aging*;31(2):161-71.
69. Ambarish V, Chandrashekara S, Suresh KP. Moderate regular exercises reduce inflammatory response for physical stress. *Indian J Physiol Pharmacol*;56(1):7-14.
70. Waldner MJ, Foersch S, Neurath MF. Interleukin-6--a key regulator of colorectal cancer development. *Int J Biol Sci*;8(9):1248-53.
71. Lee WS, Baek JH, You DH, Nam MJ. Prognostic value of circulating cytokines for stage III colon cancer. *J Surg Res*: S0022-4804(12)00796-2.
72. Baltgalvis KA, Berger FG, Pena MM, Davis JM, Muga SJ, Carson JA. Interleukin-6 and cachexia in *ApcMin/+* mice. *Am J Physiol Regul Integr Comp Physiol.*;294(2):R393-401.

73. Mehl KA, Davis JM, Clements JM, Berger FG, Pena MM, Carson JA. Decreased intestinal polyp multiplicity is related to exercise mode and gender in ApcMin/+ mice. *J Appl Physiol*;98(6):2219-25.
74. Puppa MJ, White JP, Velázquez KT, Baltgalvis KA, Sato S, Baynes JW, Carson JA. The effect of exercise on IL-6-induced cachexia in the Apc (Min/+) mouse. *J Cachexia Sarcopenia Muscle*. 2012 Jun;3(2):117-37.
75. White JP, Puppa MJ, Sato S, Gao S, Price RL, Baynes JW, Kostek MC, Matesic LE, Carson JA. IL-6 regulation on skeletal muscle mitochondrial remodeling during cancer cachexia in the ApcMin/+ mouse. *Skelet Muscle*. 2012 Jul 6;2:14. doi: 10.1186/2044-5040-2-14.
76. Bradshaw AD, Sage EH. SPARC, a matricellular protein that functions in cellular differentiation and tissue response to injury. *J Clin Invest* 2001;107:1049-54.
77. Aoi W, Naito Y, Takagi T, Tanimura Y, Takanami Y, Kawai Y, Sakuma K, Hang LP, Mizushima K, Hirai Y, Koyama R, Wada S, Higashi A, Kokura S, Ichikawa H, Yoshikawa T. A novel myokine, secreted protein acidic and rich in cysteine (SPARC), suppresses colon tumorigenesis via regular exercise. *Gut*. 2012 Nov 16.
78. World Health Organization. Global recommendations on physical activity for health 2008.
79. Fuku N, Ochiai M, Terada S, Fujimoto E, Nakagama H, Tabata I. Effect of running training on DMH-induced aberrant crypt foci in rat colon. *Med Sci Sports Exerc*. 2007 Jan;39(1):70-4.
80. Demarzo MM, Martins LV, Fernandes CR, Herrero FA, Perez SE, Turatti A, Garcia SB. Exercise reduces inflammation and cell proliferation in rat colon carcinogenesis. *Med Sci Sports Exerc*. 2008 Apr;40(4):618-21.
81. McTiernan A, Yasui Y, Sorensen B, Irwin ML, Morgan A, Rudolph RE, Surawicz C, Lampe JW, Ayub K, Potter JD, Lampe PD. Effect of a 12-month exercise intervention on patterns of cellular proliferation in colonic crypts: a randomized controlled trial. *Cancer Epidemiol Biomarkers Prev*. 2006 Sep;15(9):1588-97.
82. Bostick RM, Fosdick L, Lillemoie TJ, Overn P, Wood JR, Grambsch P, Elmer P, Potter JD. Methodological findings and considerations in measuring colorectal epithelial cell proliferation in humans. *Cancer Epidemiol Biomarkers Prev*. 1997 Nov;6(11):931-42.
83. Wolin KY, Yan Y, Colditz GA. Physical activity and risk of colon adenoma: a meta-analysis. *Br J Cancer*. 2011 Mar 1;104(5):882-5.

84. Sanchez NF, Stierman B, Saab S, Mahajan D, Yeung H, Francois F. Physical activity reduces risk for colon polyps in a multiethnic colorectal cancer screening population. BMC Res Notes. 2012 Jun 20;5:312.

BIBLIOGRAPHY

Admiraal WM, van Valkengoed IG, JS LdM, Stronks K, Hoekstra JB, Holleman F. The association of physical inactivity with Type 2 diabetes among different ethnic groups. *Diabet Med*;28:668-72.

Alibegovic AC, Sonne MP, Hojbjerg L, et al. Insulin resistance induced by physical inactivity is associated with multiple transcriptional changes in skeletal muscle in young men. *Am J Physiol Endocrinol Metab*;299:E752-63.

Ambarish V, Chandrashekara S, Suresh KP. Moderate regular exercises reduce inflammatory response for physical stress. *Indian J Physiol Pharmacol*;56(1):7-14.

Aoi W, Naito Y, Takagi T, Kokura S, Mizushima K, Takanami Y, Kawai Y, Tanimura Y, Hung LP, Koyama R, Ichikawa H, Yoshikawa T. Regular exercise reduces colon tumorigenesis associated with suppression of iNOS. *Biochem Biophys Res Commun*.;399(1):14-9.

Aoi W, Naito Y, Takagi T, Tanimura Y, Takanami Y, Kawai Y, Sakuma K, Hang LP, Mizushima K, Hirai Y, Koyama R, Wada S, Higashi A, Kokura S, Ichikawa H, Yoshikawa T. A novel myokine, secreted protein acidic and rich in cysteine (SPARC), suppresses colon tumorigenesis via regular exercise. *Gut*. 2012 Nov 16.

Aparicio T, Kotelevets L, Tsocas A, et al. Leptin stimulates the proliferation of human colon cancer cells in vitro but does not promote the growth of colon cancer xenografts in nude mice or intestinal tumorigenesis in *Apc(Min/+)* mice. *Gut* 2005;54:1136-45.

Arsenault BJ, Rana JS, Lemieux I, et al. Physical inactivity, abdominal obesity and risk of coronary heart disease in apparently healthy men and women. *Int J Obes (Lond)*;34:340-7.

Baltgalvis KA, Berger FG, Pena MM, Davis JM, Muga SJ, Carson JA. Interleukin-6 and cachexia in *ApcMin/+* mice. *Am J Physiol Regul Integr Comp Physiol*.;294(2):R393-401.

Bardou M, Barkun AN, Martel M. Obesity and colorectal cancer. *Gut*. 2013 Mar 12.

Baserga, R., Peruzzi, F. and Reiss, K. The IGF-1 receptor in cancer biology. *Int. J. Cancer* 2003, 107, 873–877.

Bostick RM, Fosdick L, Lillemoe TJ, Overn P, Wood JR, Grambsch P, Elmer P, Potter JD. Methodological findings and considerations in measuring colorectal epithelial cell proliferation in humans. *Cancer Epidemiol Biomarkers Prev.* 1997 Nov;6(11):931-42.

Bradshaw AD, Sage EH. SPARC, a matricellular protein that functions in cellular differentiation and tissue response to injury. *J Clin Invest* 2001;107:1049-54.

Carnethon MR. Physical Activity and Cardiovascular Disease: How Much is Enough? *Am J Lifestyle Med* 2009;3:44S-9S.

Chi F, Wu R, Zeng YC, Xing R, Liu Y. Circulation insulin-like growth factor peptides and colorectal cancer risk: an updated systematic review and meta-analysis. *Mol Biol Rep.* 2013 May;40(5):3583-90.

Colbert LH, Mai V, Perkins SN, Berrigan D, Lavigne JA, Wimbrow HH, Alvord WG, Haines DC, Srinivas P, Hursting SD. Exercise and intestinal polyp development in APCMin mice. *Med Sci Sports Exerc.* 2003 Oct;35(10):1662-9.

Colbert LH, Mai V, Tooze JA, Perkins SN, Berrigan D, Hursting SD. Negative energy balance induced by voluntary wheel running inhibits polyp development in APCMin mice. *Carcinogenesis.* 2006 Oct;27(10):2103-7.

Corpet DE, Jacquinet C, Peiffer G, et al. Insulin injections promote the growth of aberrant crypt foci in the colon of rats. *Nutr Cancer*;27:316–320.

DeFronzo RA, Ferrannini E. Insulin resistance: a multifaceted syndrome responsible for NIDDM, obesity, hypertension, dyslipidemia, and atherosclerotic cardiovascular disease. *Diabetes Care* ;14:173–194.

Demarzo MM, Martins LV, Fernandes CR, Herrero FA, Perez SE, Turatti A, Garcia SB. Exercise reduces inflammation and cell proliferation in rat colon carcinogenesis. *Med Sci Sports Exerc.* 2008 Apr;40(4):618-21.

Eckardt K, Taube A, Eckel J. Obesity-associated insulin resistance in skeletal muscle: role of lipid accumulation and physical inactivity. *Rev Endocr Metab Disord*;12:163-72.

Endo H, Hosono K, Uchiyama T, et al. Leptin acts as a growth factor for colorectal tumours at stages subsequent to tumour initiation in murine colon carcinogenesis. *Gut*;60:1363-71.

Fenton JI, Birmingham JM, Hursting SD, Hord NG. Adiponectin blocks multiple signaling cascades associated with leptin-induced cell proliferation in Apc Min/+ colon epithelial cells. *Int J Cancer*;122(11):2437-45.

Fenton JI, Hursting SD, Perkins SN, Hord NG. Leptin induces an Apc genotype-associated colon epithelial cell chemokine production pattern associated with macrophage chemotaxis and activation. *Carcinogenesis* 2007;28:455-64.

Friedenreich CM, Neilson HK, Lynch BM. State of the epidemiological evidence on physical activity and cancer prevention. *Eur J Cancer*;46:2593-604.

Fuku N, Ochiai M, Terada S, Fujimoto E, Nakagama H, Tabata I. Effect of running training on DMH-induced aberrant crypt foci in rat colon. *Med Sci Sports Exerc.*2007Jan;39(1):70-4.

Gialamas SP, Petridou ET, Tseleni-Balafouta S, Spyridopoulos TN, Matsoukis IL, Kondi-Pafiti A, Zografos G, Mantzoros CS. Serum adiponectin levels and tissue expression of adiponectin receptors are associated with risk, stage, and grade of colorectal cancer. *Metabolism.*;60(11):1530-8.

Giovannucci E. Insulin and colon cancer. *Cancer Causes Control*;6:164–179.

González-Sánchez JL, Zabena CA, Martínez-Larrad MT, Fernández-Pérez C, Pérez-Barba M, Laakso M, Serrano-Ríos M. An SNP in the adiponectin gene is associated with decreased serum adiponectin levels and risk for impaired glucose tolerance. *Obes Res*;13(5):807-12.

Greten FR, Eckmann L, Greten TF, Park JM, Li ZW, Egan LJ, Kagnoff MF, Karin M. IKKbeta links inflammation and tumorigenesis in a mouse model of colitis-associated cancer. *Cell*;118(3):285-96.

Guo YS, Narayan S, Yallampalli C, et al. Characterization of insulin like growth factor I receptors in human colon cancer. *Gastroenterology.* 1992;102:1101–1108.

Harriss DJ, Cable NT, George K, Reilly T, Renehan AG, Haboubi N. Physical activity before and after diagnosis of colorectal cancer: disease risk, clinical outcomes, response pathways and biomarkers. *Sports Med*;37(11):947-60.

Hoffman-Goetz L, Pervaiz N, Guan J. Voluntary exercise training in mice increases the expression of antioxidant enzymes and decreases the expression of TNF-alpha in intestinal lymphocytes *Brain Behav Immun*;23(4):498-506.

Hojbjerre L, Sonne MP, Alibegovic AC, et al. Impact of physical inactivity on subcutaneous adipose tissue metabolism in healthy young male offspring of patients with type 2 diabetes. *Diabetes*;59:2790-8.

Joshu CE, Parmigiani G, Colditz G, Platz EA. Opportunities for the primary prevention of colorectal cancer in the United States. *Cancer Prev Res (Phila)*;1:138-45.

Ju J, Nolan B, Cheh M, Bose M, Lin Y, et al. Voluntary exercise inhibits intestinal tumorigenesis in ApcMin/+ mice and azoxymethane/dextran sulfate sodium-treated mice. *BMC Cancer*. 2008;8: 316.

Kadoglou NP, Fotiadis G, Athanasiadou Z, Vitta I, Lampropoulos S, Vrabas IS. The effects of resistance training on ApoB/ApoA-I ratio, Lp(a) and inflammatory markers in patients with type 2 diabetes. *Endocrine*;42(3):561-9.

Khalili H, Chan AT. Is diabetes a risk factor for colorectal cancer? *Dig Dis Sci*;57(6):1427-9.

Kim YI. Diet, lifestyle, and colorectal cancer: is hyperinsulinemia the missing link. *Nutr Rev*;56:275–279.

Koenuma M, Yamori T, Tsuruo T (1989) Insulin and insulin-like growth factor 1 stimulate proliferation of metastatic variants of colon carcinoma 26. *Jpn J Cancer Res* 80:51–58.

Kokkinos P, Sheriff H, Kheirbek R. Physical inactivity and mortality risk. *Cardiol Res Pract*;2011:924945.

Koohestani N, Tran TT, Lee W, et al. Insulin resistance and promotion of aberrant crypt foci in the colons of rats on a high fat diet. *Nutr Cancer*;29:69–76.

Lee KY, Kang HS, Shin YA. Exercise improves adiponectin concentrations irrespective of the adiponectin gene polymorphisms SNP45 and the SNP276 in obese Korean women. *Gene*;516(2):271-6.

Lee WS, Baek JH, You DH, Nam MJ. Prognostic value of circulating cytokines for stage III colon cancer. *J Surg Res*: S0022-4804(12)00796-2.

Lefrançois L, Lycke N. Isolation of mouse small intestinal intraepithelial lymphocytes, Peyer's patch, and lamina propria cells. *Clin Chem*; 52(1):97-103.

Ma J, Pollak MN, Giovannucci E, Chan JM, Tao Y, Hennekens CH, Stampfer MJ. Prospective study of colorectal cancer risk in men and plasma levels of insulin-like growth factor (IGF)-I and IGF-binding protein-3. *J Natl Cancer Inst*.1999Apr 7;91(7):620-5.

Manousos O, Souglakos J, Bosetti C, Tzonou A, Chatzidakis V, Trichopoulos D, Adami HO, Mantzoros C. IGF-I and IGF-II in relation to colorectal cancer. *Int J Cancer*. 1999 Sep 24;83(1):15-7.

Mariusz Koda, Mariola Sulkowska, Luiza Kanczuga-Koda, Eva Surmacz, and Stanislaw Sulkowski. Overexpression of the obesity hormone leptin in human colorectal cancer. *J Clin Pathol*.2007 August;60(8): 902–906.

McClellan JL, Davis JM, Steiner JL, Day SD, Steck SE, Carmichael MD, Murphy EA. Intestinal inflammatory cytokine response in relation to tumorigenesis in the Apc(Min/+) mouse. *Cytokine*;57(1):113-9.

McClellan JL, Davis JM, Steiner JL, Enos RT, Jung SH, Carson JA, Pena MM, Carnevale KA, Berger FG, Murphy EA. Linking tumor-associated macrophages, inflammation, and intestinal tumorigenesis: role of MCP-1. *Am J Physiol Gastrointest Liver Physiol*;303(10):G1087-95.

McTiernan A, Yasui Y, Sorensen B, Irwin ML, Morgan A, Rudolph RE, Surawicz C, Lampe JW, Ayub K, Potter JD, Lampe PD. Effect of a 12-month exercise intervention on patterns of cellular proliferation in colonic crypts: a randomized controlled trial. *Cancer Epidemiol Biomarkers Prev*. 2006 Sep;15(9):1588-97.

Mehl KA, Davis JM, Clements JM, Berger FG, Pena MM, Carson JA. Decreased intestinal polyp multiplicity is related to exercise mode and gender in ApcMin/+ mice. *J Appl Physiol*;98(6):2219-25.

Mendis S. The contribution of the Framingham Heart Study to the prevention of cardiovascular disease: a global perspective. *Prog Cardiovasc Dis*;53:10-4.

Mutoh M, Teraoka N, Takasu S, Takahashi M, et al. Loss of Adiponectin Promotes Intestinal Carcinogenesis in Min and Wild-type Mice. *Gastroenterology*. 2011 Jun;140(7):2000-8.

Nahleh Z, Bhatti NS, Mal M. How to reduce your cancer risk: mechanisms and myths. *Int J Gen Med*;4:277-87.

Naylor MS, Stamp GW, Balkwill FR. Investigation of cytokine gene expression in human colorectal cancer. *Cancer Res*; 50 (14):4436–4440.

Onizawa M, Nagaishi T, Kanai T, Nagano K, Oshima S, Nemoto Y, Yoshioka A, Totsuka T, Okamoto R, Nakamura T, Sakamoto N, Tsuchiya K, Aoki K, Ohya K, Yagita H, Watanabe H. Signaling pathway via TNF-alpha/NF-kappaB in intestinal epithelial cells may be directly involved in colitis-associated carcinogenesis. *Am. J. Physiol. Gastrointest. Liver Physiol.*;296 (4):G850–859.

Otani K, Kitayama J, Yasuda K, Nio Y, Iwabu M, Okudaira S, Aoki J, Yamauchi T, Kadowaki T, Nagawa H. Adiponectin suppresses tumorigenesis in Apc(Min)(/+) mice. *Cancer Lett*. 2010 Feb 28;288(2):177-82.

Ott JJ, Ullrich A, Mascarenhas M, Stevens GA. Global cancer incidence and mortality caused by behavior and infection. *J Public Health (Oxf)*;33:223-33.

Packer N, Hoffman-Goetz L. Exercise training reduces inflammatory mediators in the intestinal tract of healthy older adult mice. *Can J Aging*;31(2):161-71.

Pollak M. Insulin and insulin-like growth factor signaling in neoplasia. *Nat Rev Cancer*. 2008;8:915–928.

Pollak M. Insulin, insulin-like growth factors and neoplasia. *Best Pract Res Clin Endocrinol Metab*. 2008;22:625–638.

Powrie F. Immune regulation in the intestine: a balancing act between effector and regulatory T cell responses. *Ann N Y Acad Sci*;1029:132-41.

Puppa MJ, White JP, Velázquez KT, Baltgalvis KA, Sato S, Baynes JW, Carson JA. The effect of exercise on IL-6-induced cachexia in the Apc (Min/+) mouse. *J Cachexia Sarcopenia Muscle*. 2012 Jun;3(2):117-37.

Rustgi AK. The genetics of hereditary colon cancer. *Genes Dev* 2007;21:2525-38.

Sanchez NF, Stierman B, Saab S, Mahajan D, Yeung H, Francois F. Physical activity reduces risk for colon polyps in a multiethnic colorectal cancer screening population. *BMC Res Notes*. 2012 Jun 20;5:312.

Saxena A, Baliga M, et al. Mucus and adiponectin deficiency: role in chronic inflammation-induced colon cancer. *Int J Colorectal Dis*. 2013 Mar 9.

Saxena A, Chumanevich A, Fletcher E, Larsen B, Lattwein K, Kaur K, Fayad R. Adiponectin deficiency: role in chronic inflammation induced colon cancer. *Biochim Biophys Acta*. 2012 Apr;1822(4):527-36.

Saxena A, Fletcher E, Larsen B, Baliga MS, Durstine JL, Fayad R. Effect of exercise on chemically-induced colitis in adiponectin deficient mice. *J Inflamm (Lond)*. 2012 Aug 21;9(1):30.

Schwandt P, Liepold E, Bertsch T, Haas GM. Lifestyle, Cardiovascular Drugs and Risk Factors in Younger and Elder Adults: The PEP Family Heart Study. *Int J Prev Med*;1:56-61.

Sikalidis AK, Varamini B. Roles of hormones and signaling molecules in describing the relationship between obesity and colon cancer. *Pathol Oncol Res*. 2011 Dec;17(4):785-90.

Singh P, Guo YS, Narayan S, et al. IGF-I and IGF-I receptor in mouse colon. *In Vitro Cell Dev Biol*;27A(10):755-8.

Tang H, Xie M, et al. The roles of aerobic exercise training and suppression IL-6 gene expression by RNA interference in the development of insulin resistance. *Cytokine*.;61(2):394-405.

Terzić J, Grivennikov S, Karin E, Karin M. Inflammation and colon cancer. *Gastroenterology*;138(6):2101-2114.

Thyfault JP, Booth FW. Lack of regular physical exercise or too much inactivity. *Curr Opin Clin Nutr Metab Care*;14:374-8.

Waldner MJ, Foersch S, Neurath MF. Interleukin-6--a key regulator of colorectal cancer development. *Int J Biol Sci*;8(9):1248-53.

White JP, Puppa MJ, Sato S, Gao S, Price RL, Baynes JW, Kostek MC, Matesic LE, Carson JA. IL-6 regulation on skeletal muscle mitochondrial remodeling during cancer cachexia in the *ApcMin/+* mouse. *Skelet Muscle*. 2012 Jul 6;2:14. doi: 10.1186/2044-5040-2-14.

Wiggins MS, Simonavice EM. Cancer prevention, aerobic capacity, and physical functioning in survivors related to physical activity: a recent review. *Cancer Manag Res*;2:157-64.

Wilson JA. Tumor necrosis factor alpha and colitis-associated colon cancer. *N Engl J Med*;358(25):2733-4.

Wise FM. Coronary heart disease--the benefits of exercise. *Aust Fam Physician*;39:129-33.

Wolin KY, Yan Y, Colditz GA. Physical activity and risk of colon adenoma: a meta-analysis. *Br J Cancer*. 2011 Mar 1;104(5):882-5.

Woo SK, Kang HS. Apolipoprotein C-III SstI genotypes modulate exercise-induced hypotriglyceridemia. *Med Sci Sports Exerc*. 2004 Jun;36(6):955-9.

World Health Organization. Global recommendations on physical activity for health 2008.

Wu, Y. P., Yakar, S., Zhao, L., Hennighausen, L. and LeRoith, D. Circulating insulin-like growth factor-I levels regulate colon cancer growth and metastasis. *Cancer* 2002, Res. 62,1030–1035.

Yoon J, Son K, Eom C, Durrance D, Park S. Pre-existing diabetes mellitus increases the risk of gastric cancer: A meta-analysis. *World J Gastroenterol*; 19(6):936-945.